

1 Macrophyte presence and growth form influence macroinvertebrate community structure

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3 Peter D. Walker<sup>1,4\*</sup>, Sander Wijnhoven<sup>2</sup> and Gerard van der Velde<sup>1,3</sup>

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5 <sup>1</sup>*Department of Animal Ecology and Ecophysiology, Institute for Water and Wetland Research, Faculty of*  
6 *Science, Radboud University Nijmegen, Nijmegen, The Netherlands*

7 <sup>2</sup>*Monitor Taskforce, Royal Netherlands Institute for Sea Research, NIOZ-Yerseke, Yerseke, The*  
8 *Netherlands*

9 <sup>3</sup>*Netherlands Centre for Biodiversity Naturalis, Leiden, The Netherlands*

10 <sup>4</sup>*APEM Ltd, Centre for Innovation and Enterprise, Oxford University Begbroke Science Park, Oxfordshire,*  
11 *UK*

12  
13  
14 \*Corresponding author at: APEM Ltd, Centre for Innovation and Enterprise, Oxford University Begbroke  
15 Science Park, Sandy Lane, Yarnton, Oxfordshire OX5 1PF, UK.

16 E-mail: [p.walker@apemltd.co.uk](mailto:p.walker@apemltd.co.uk); Tel: +44 (0)1865 854853; Fax: +44 (0)1865 854801.

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2 ABSTRACT

3 Multivariate analysis demonstrated that macroinvertebrate assemblages of macrophyte-dominated sub-  
4 habitats within a small eutrophic pond differed markedly from those of *Bottom substrate* and *Open water*  
5 habitats. Certain habitats (e.g. *Nymphaea* and *Phragmites*) appeared to be quite similar in their  
6 macroinvertebrate communities, whereas others appeared to be very distinct in terms of the species  
7 composition (e.g. *Open water* habitat). Analysis of functional feeding groups also revealed differences  
8 between habitats in terms of the community structure. Again, the *Open water* habitat exhibiting the most  
9 marked difference.

10 Macrophyte growth form does not cause significant differences in macroinvertebrate species richness and  
11 diversity but it has a significant effect on macroinvertebrate abundance. Habitats consisting of highly  
12 branched and dissected macrophyte growth forms provide more food resources and microhabitats  
13 supporting larger numbers of macroinvertebrates than macrophytes with firm undissected stalks and leaves.

14 This study highlights the importance of maintaining the ecological quality of small freshwater habitats in  
15 order to promote macrophyte growth and thus maintain a high level of species richness within such  
16 ecosystems.

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18 *Key words:* macroinvertebrates, macrophytes, growth form, communities, pond

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## 1. Introduction

The physical structure and architectural complexity of freshwater habitats determine the community that resides there (Rennie and Jackson 2005). Macroinvertebrate distribution patterns in streams are typically dependent upon the particle size of the mineral component of the substrate (e.g. Tolkamp and Both, 1978; Khalaf and Tachet, 1980; Sheldon and Haick, 1981). Within still water bodies (e.g. lakes and ponds) it has been demonstrated that macroinvertebrate distribution, is predominantly determined by the vegetation type, and more importantly, by the architecture or growth form of the dominant macrophytes (e.g. Cyr and Downing, 1988; Duggan et al., 2001; Dvorak and Best, 1982; Karassowska and Mikulski, 1960; Korinkova, 1971; Rennie and Jackson, 2005; Scheffer et al., 1984; Taniguchi et al., 2003).

The importance of aquatic macrophytes as habitats in aquatic systems has been demonstrated by the abundant and diverse macroinvertebrate communities that they support (Warfe and Barmuta, 2004). Both macroinvertebrate diversity and abundance increase with increasing vegetation biomass and/or density (Warfe and Barmuta, 2004; and references therein). Macrophytes fill the water column in a characteristic way providing extensive substrate for periphyton, macroinvertebrates and developing eggs, as well as shelter against predation by larger animals such as fish. Furthermore, macrophytes influence the under water climate and chemistry via their uptake and release of chemical substances (e.g. nutrients, allelopathic substances) and influence bottom life via the input of macrophyte derived detritus (Van der Valk, 2006). Macrophytes therefore structure lentic communities (Den Hartog and Van der Velde, 1988; Jeppesen et al., 1998). Macrophyte growth form, adding substrate and biomass with a particular architecture and structural complexity, has been referred as a driving factor behind macroinvertebrate community composition (Den Hartog and Van der Velde, 1988; Jeffries, 1993; Van der Velde, 1980).

In this study the macroinvertebrate communities of several sub-habitats, represented by various macrophyte growth forms, within a single pond are described. Studying the influence of various growth forms of macrophytes within a single pond has the advantage that water chemistry and physical conditions are similar within the various types of sub-habitat and show comparable fluctuations, while macroinvertebrates represent one species pool and can easily select where to reside. Seasonal variation should normally be taken into consideration but can be ruled out here because all samples were obtained

1 during the same period. In this way the direct influence of growth form on macroinvertebrate communities  
2 can be studied without large variation in other factors.

3 In this paper the macroinvertebrate species richness, diversity and abundance are described from seven  
4 different sub-habitats (five different macrophyte growth forms, bare bottom substrate and open water)  
5 within a freshwater pond. This study thus provides useful information pertaining to the ecological  
6 importance of macrophytes in small freshwater systems.

## 8 **2. Study area**

9 This investigation was conducted in a small, eutrophic, still water pond in Fleetwood, NW England  
10 (Ordnance Survey Grid Reference: SD318449) (open water surface area approximately 700 m<sup>2</sup>), with an  
11 average depth of <100 cm and a maximum depth of 140 cm. The bottom substrate is clay overlain with  
12 coarse gravel and fine organic silt.

13 Encroachment of vegetation is clearly evident from a large reed bed consisting predominantly of  
14 *Phragmites australis* interspersed with *Typha latifolia* and *Iris pseudacorus* at the southern end of the pond  
15 which is approximately the same size as the open pond itself. Almost the entire littoral zone is vegetated  
16 with five macrophyte species belonging to different growth forms predominating (Table 1).

## 18 **3. Materials and Methods**

### 19 *3.1. Field collections*

20 A standard pond-net (230 x 255 mm frame with 900 µm mesh) was used to collect 10 replicate  
21 macroinvertebrate samples from each of the seven main habitat types identified within the pond. Care was  
22 taken to avoid repeat sampling of the same area. Each sample was obtained using 1m sweeps with the net  
23 being covered at the end of each sweep to prevent escape or contamination of the sample as the net was  
24 removed from the sampling point. This ensured that a standard volume of approximately 50,000 cm<sup>3</sup> was  
25 sampled each time. Samples taken from the bottom substrate were obtained by pushing the net rim  
26 approximately 2 cm into the substrate and then carrying out a 1m sweep. For macrophyte habitats, samples  
27 were obtained from stands representing several growth forms as close to monospecific as possible. Care  
28 was taken to ensure that each net sweep was performed only in that specific habitat. i.e. when sampling

weed beds care was taken not to touch, or disturb, the bottom sediment to avoid sampling invertebrate populations residing there. Similarly care was taken not to sample open water habitats above or surrounding weed beds to avoid sampling invertebrates in that region. All sampling was conducted within a 1 week period during July to avoid any potential differences associated with season. Whilst this study therefore pertains to only a limited time, this month is associated with high productivity for both macrophytes and macroinvertebrates thus main lines are likely to be observed.

Samples were immediately rinsed into 1 litre plastic containers using de-ionised water and the net was thoroughly examined for any invertebrates still clinging to the net. Any macroinvertebrates found were gently removed using forceps or de-ionised water and added to the sample. Samples were later sorted in large white trays and all macro-invertebrates were removed from the sediment, water and/or vegetation, and preserved in 70% ethanol. Macroinvertebrates were identified to species level where possible.

### 3.2. Statistical methods

Berger-Parker Dominance Index (BPDI) was used as a simple measure of species diversity and is calculated as:

BPDI score = Number of individuals of the most abundant species/Total number of Individuals of the sample

BPDI scores closer to 0 indicate higher species diversity.

Data for macroinvertebrate abundance, species richness and species diversity for each habitat were tested for significant differences between habitats using a Kruskal-Wallis ANOVA followed by Dunns multiple comparisons post hoc test. Spearmans rank correlation coefficients were performed to test for linear relationships between these variables.

### 3.3. Multivariate analyses

A Canonical Correspondence Analysis (CCA), which is a direct gradient analysis, was performed on the log-transformed species data, using the CANOCO for Windows software package (version 4.5) (Ter Braak and Smilauer 1998). A Detrended Correspondence Analysis (DCA) showed that the data had a long gradient length (4.1 for all data; 2.2 when open water data were excluded); therefore a unimodal ordination

method was used. A CCA is a direct method, which means that in this case the species compositions can be directly explained by the environmental characteristics (habitat types).

To compare (dis)similarity of the communities between habitat types non-metric multi-dimensional scaling (nMDS) in combination with analyses of similarity (ANOSIM) were executed in Primer 5.2.9. Before analyses, all data were fourth-root transformed to minimize the effect of dominant species, and similarity analyses are based on the Bray-Curtis formula (Clarke and Gorley, 2001). An nMDS 2D representation was considered acceptable when the stress factor did not transgress 0.2. With ANOSIM, pair-wise comparisons of differences between habitat-types in the macrofauna communities were tested for taking a Bonferroni correction for multiple testing of the same kind, according to  $p \leq 0.05/N$  ( $N$  = number of tests of the same kind) into account. One sample from the 'Open water' was excluded from the analyses as it contained no macrofauna.

## 4. Results

### 4.1. Growth forms

Seven key habitat types were identified (Table 1). These habitats can be further grouped into open water, bare bottom substrate, emergent helophyte (*P. australis*), nymphaeid (*Nymphaea alba*) and peplid (*Callitriche* sp.) both possessing floating leaves, and fully submerged elodeid (*Elodea canadensis*) and ceratophyllid (*Ceratophyllum demersum*) macrophytes.

The different habitat types represent a diverse range of habitats differing markedly in terms of their structural complexity. The *Open water* habitat is structurally non-complex due to the lack of vegetation or other three-dimensional components. The emergent (*Phragmites*) and floating leaved (*Nymphaea*) habitats exhibit some structural complexity with plant stalks running upwards through the water column to the water's surface. The three submerged (either partially or completely) habitat types containing macrophytes (*Elodea*, *Ceratophyllum* and *Callitriche*) all complex in terms of their structural architecture due to the highly branched and dissected growth forms they exhibit. The non-vegetated bottom (*Bottom substrate*) habitat is unique in that it is complex in terms of its structure (many and varied microhabitats) but does not project into the water column and contains no large, living, structural components.

#### 4.2. Species richness, diversity and abundance

In total 39 different taxa were identified from 2,707 individuals collected from seven different sub-habitats. The most abundant taxa overall were the water hog louse, *Asellus aquaticus* and the triclad flatworm, *Dugesia lugubris*, with 773 and 887 individuals respectively, from 70 samples. Damselfly larvae (Zygoptera; *Enallagma cyathigerum* and *Ischnura elegans*) were also common in occurrence and were the second and third most abundant predatory species (*D. lugubris* being the most abundant). Table 2 shows the occurrence and total numbers of the different taxa recorded from each habitat type.

The species richness of each habitat increased with increasing number of samples taken (Fig. 1). However, in all cases, except the *Phragmites* habitat, this number levelled off after approximately six samples had been analysed (Fig. 1). With regard to the total species richness observed, the habitats displayed the following order: *Bottom substrate*  $\geq$  *Phragmites* > *Nymphaea* > *Callitriche*  $\geq$  *Elodea* > *Ceratophyllum* > *Open water*. The *Phragmites* and *Bottom substrate* habitats were the most species rich, each found to match the species richness of the aggregated samples (for the whole pond) by > 60%. Both of these sites also contained the highest number of species unique to those habitats (three each, Table 3). The *Open water* habitat was the poorest in terms of species richness with only nine different taxa being recorded.

Kruskall-Wallis test demonstrated significant differences in species richness between the different habitats ( $P < 0.0001$ ; Table 3). Dunn's multiple comparison test revealed that values for the *open water* habitat differed significantly from those of *Bottom substrate*, *Nymphaea*, *Callitriche* and *Elodea* habitats. With regard to the mean number of species observed, the habitats displayed the following, decreasing order: *Bottom substrate* > *Callitriche* > *Elodea* > *Nymphaea*, *Ceratophyllum*, *Phragmites* > *Open water*. Differences were also shown between habitats for species diversity (Kruskall-Wallis test -  $P = 0.001$ ; Table 3) with Dunn's multiple comparison test demonstrating that these differences were only statistically significant between *Nymphaea* and *Elodea* habitats.

The total number of invertebrates collected from each habitat type showed a large range from just 19 individuals collected from *Open water* samples to 789 individuals collected from *Callitriche* samples. The total number of invertebrates collected were in decreasing order *Ceratophyllum* > *Elodea* > *Bottom substrate* > *Callitriche* > *Phragmites* > *Nymphaea* > *Open water*. Significant differences in the mean

invertebrate abundance per sample were observed between habitats (Kruskal-Wallis test -  $P < 0.0001$ ; Table 3) with Dunn's multiple comparison test demonstrating that values for *Open water* habitat were significantly different from all other habitats apart from *Phragmites* and *Nymphaea* habitats. Significant differences were also observed between *Bottom substrate* and *Nymphaea*; *Phragmites* and *Elodea*; *Phragmites* and *Ceratophyllum*; *Nymphaea* and *Elodea*; and *Nymphaea* and *Ceratophyllum*. With regard to the mean number of individuals observed, the habitats displayed the following, decreasing order: *Ceratophyllum* > *Elodea* > *Bottom substrate* > *Callitriche*, *Phragmites* > *Nymphaea* > *Open water*.

The habitats supporting the greatest number of macroinvertebrates were the *Ceratophyllum* and *Elodea* habitats. The combined abundance of these two habitats made up 56.9% of the abundance from the aggregated samples. In both cases, one taxon (*Dugesia lugubris*) constituted approximately half of the habitats overall abundance. This indicates low evenness and this is reflected by their relatively poor scores for species diversity (Table 3). The *Phragmites* habitat had the BPD I score closest to 0 and therefore exhibited the highest species diversity out of the seven habitats sampled (Table 3). BPD I score was in decreasing order *Open water* > *Elodea* > *Ceratophyllum* > *Callitriche*, *Phragmites* > *Bottom substrate* > *Nymphaea*. Differences between habitats with regard to species diversity were only found to be significant between *Nymphaea* and *Elodea* habitats (Dunn's multiple comparison test:  $P < 0.05$ ).

#### 4.3. Multivariate analyses

For the Canonical Correspondence Analysis with all data included, the *Open water* habitat appeared to be completely different from the other habitats in terms of macroinvertebrate species present (see insert, Figure 2). *Argulus foliaceus* (no. 21 in Figure 2) was found more frequently in *Open water* than in any of the other habitats; *Dryops* sp. (no. 11 in Figure 2) was observed only once. All the other taxa were observed in at least two habitats and/or in similar numbers. When the *Open water* data were excluded, the distribution of the taxa in the other habitats is more distinct (Figure 2). Eigenvalues for axes 1 to 4 are 0.295, 0.221, 0.160 and 0.101 respectively. The species – environment correlations are high, being 0.956 and 0.901 for the axes 1 and 2, respectively. The first ordination axis might be interpreted as the gradient from without vegetation to dense vegetation whereas the second axis might be related to the three-dimensional complexity of the vegetation structure in the water column. Therefore the *Nymphaea* and



*Phragmites* habitats appeared to have relatively similar species compositions. The three fully submerged, highly branched/dissected macrophyte sub-habitats also show a large degree of similarity in terms of their species composition. Few species are specifically related to *Callitriche*; these species can also be found in the other habitats in similar numbers. These findings are confirmed by the results of the analyses of similarity (Table 4). As indicated by an overall R-value of 0.517 the communities of the different habitat types are clearly distinguishable, showing significant differences ( $p=0.001$ ). The nMDS plot of Figure 3 indicates that there is some overlap in the species compositions but the samples for each of the habitat types cluster very well. Pair-wise comparisons (ANOSIM results Table 4) show that actually the communities of each of the habitat types significantly differ from each other habitat type, except for the mutual communities related to *Callitriche* and *Phragmites*, and the mutual communities of *Elodea* and *Ceratophyllum* habitat.

#### 4.4. Functional Feeding Groups

Analysis of functional feeding groups showed that the overall abundance of predatory carnivores was higher than all other groups (Figure 4). The group containing shredders, grazers and scrapers was the second largest of the five groups and the parasitic carnivores constituted only 1% of the pond community overall. This pattern is similar for macrophyte-dominated habitats (*Phragmites*, *Callitriche*, *Ceratophyllum*, *Nymphaea* and *Elodea* (Figure 4)). The *Bottom substrate* and *Phragmites* habitat contained relatively most filter feeders and detritivores of all habitats studied. In the *Bottom substrate* habitat, predatory carnivores are only outnumbered by the group containing the shredders, grazers and scrapers. Only in the *Open water* habitat do predatory carnivores occur in fewer numbers than two of the other functional feeding groups. Parasitic carnivores were not found in large numbers although they did constitute the dominant (in terms of total number of individuals) functional feeding group in the *Open water* habitat.

### 5. Discussion

Zonation of aquatic macrophytes is typical in still-water-bodies and was also evident in the system studied here. This enabled us to perform sampling in relatively monotypic macrophyte stands. Some clear patterns in the macroinvertebrate communities of different macrophyte stands were apparent. The

1 rarefaction curves show that six samples are sufficient to estimate the species richness in each of the  
2 habitats (with the possible exception of the *Phragmites* habitat). In addition, the fact that only one operator  
3 collected and sorted all the samples reduces the margin for error that can occur when the same sampling  
4 technique is interpreted and employed by different researchers (Furse et al. 1981). Pond-net sampling is not  
5 ideal for recording quantitative data but Humphries et al. (1998) demonstrated that it is the most  
6 appropriate method when species lists are required.

7 Structurally more complex habitats are generally assumed to be richer in the number of taxa residing  
8 there due to the greater range of microhabitats offering a greater range of niches. Kreeker (1939) and  
9 Andrews and Hasler (1943) found that generally, the greater the leaf dissection of a submerged  
10 macrophyte, the larger and usually more varied was the animal population associated with it (Rosine,  
11 1955). In addition, research has previously demonstrated that macrophyte species with a higher level of  
12 structural complexity (i.e. finely dissected leaf structure and intricate branching) seem to support a greater  
13 number of individual macroinvertebrates and a greater array of different taxa (Heck and Orth, 1980; Rooke,  
14 1986). Jeffries (1993) predicted that the abundance of taxa and individuals should increase with increasing  
15 fractal complexity, which was demonstrated in experiments with artificial pondweeds of differing fractal  
16 dimension. Our data support the hypothesis that macrophytes with greater degrees of branching and leaf  
17 dissection support a greater number of macroinvertebrates but not necessarily a greater range of taxa.

18 In those habitats where relative abundance was highest (i.e. *Callitriche*, *Ceratophyllum*, *Elodea* and  
19 *Bottom substrate*), it was noted that one species typically dominated the samples. In particular *Asellus*  
20 *aquaticus* and *Dugesia lugubris* occurred in high numbers in several habitats. *A. aquaticus* is known to be  
21 relatively non-specific, in terms of its diet, feeding on detritus, periphyton and even decaying macrophyte  
22 tissues (Soszka, 1975) or in the case of *Elodea* also young leaves (Marcus et al., 1978). *A. aquaticus* is not  
23 able to feed on living leaves of *Nymphaea*, because of high phenolic content but can feed on decaying  
24 leaves of that plant (Kok et al., 1992). *Elodea* contains low amounts of phenolic compounds (Smolders et  
25 al., 2000). *D. lugubris* is a predatory species but its prey species are typically small, relatively numerous  
26 invertebrates found associated with macrophyte surfaces. This species abundance was highest on the three  
27 fully submerged macrophyte species which also have the most branched and dissected growth forms and

1 therefore can be considered the most complex. Their greater surface area provides a much larger  
2 colonisable surface for the prey items of *D. lugubris* in particular snails and *Asellus*.

3 *Open water* samples were significantly poorer than all other habitats in terms of the total number of  
4 species, species diversity and invertebrate abundance. Few macroinvertebrates are specialist pelagic  
5 feeders. Many smaller invertebrate species (e.g. some cladocerans and copepod species) are able to filter  
6 feed in this habitat type however the large number of resident fish (*personal observations*) will undoubtedly  
7 impact heavily upon any invertebrates, both macro and smaller, residing in this sub-habitat. *Argulus*  
8 *foliaceus* is an intermittent crustacean ectoparasite on fish. It has been shown to employ two host-searching  
9 strategies, one in dark conditions and the other during light periods. During light periods this species  
10 employs a 'sit-and-wait' strategy, hovering in the water column waiting for a potential host fish to swim  
11 past so that it can 'leap' on to it (Mikheev et al., 2000). This behaviour offers a logical explanation for this  
12 species presence in *Open water* samples.

13 The SGS species were frequently found in several habitats. This is probably linked to the generalist  
14 feeding strategies typical of these species. It is clear from the CANOCO analysis that they prefer to reside  
15 in the fully submerged macrophyte stands. These macrophyte species are highly dissected and branched in  
16 structure presenting a relatively high surface area for periphyton to colonize and also for detritus falling  
17 through the water column to settle upon. This provides a proportionately abundant food resource for  
18 generalist feeders such as *A. aquaticus*. In addition, high numbers of very small invertebrates such as  
19 rotifers, gastrotrichs, copepods, cladocerans and very young individuals of other common species (e.g.  
20 gastropods) as well as *Asellus* are likely to be found on these surfaces providing an abundant prey source  
21 for some of the generalist predators such as *D. lugubris* (Reynoldson and Young, 1963) and the zygopteran  
22 species.

23 The large numbers of predatory carnivores suggests that smaller prey organisms must also be present in  
24 relatively high numbers. However, large numbers of potential prey species were not generally observed  
25 apart from *A. aquaticus*. We speculate that these predatory species are predating upon smaller crustaceans  
26 such as cladocerans and copepods which were not recorded in this study probably due to the fact that most  
27 would pass through the mesh of the net used. It is also probable that these predatory species show a distinct  
28 lack of specificity with regard to what they would catch and eat.

1 In summary, the CCA analyses suggest that although seven habitat types were studied, they can be  
2 roughly grouped into four categories with the *Phragmites* (helophyte) and *Nymphaea* (nymphaeid) forming  
3 one group; *Callitriche* (peplid), *Ceratophyllum* (ceratophyllid) and *Elodea* (elodeid) forming a second  
4 group (although it should be stated that the *Callitriche* habitat is not as easily separated out as the others),  
5 the *Bottom substrate* forming a third group and finally the *Open water* habitat forming a very distinctive  
6 fourth group on its own. The nMDS results (Figure 3) confirm the distinction of these community types  
7 although each vegetation type also clearly holds its own community. It is particularly the larger variation in  
8 community assemblage for a vegetation type like *Phragmites* or *Ceratophyllum* that leads to some overlap  
9 with the communities of other vegetation types, than that their communities would not have unique  
10 elements. The more gradient-like community change from *Ceratophyllum* via *Elodea* to *Callitriche*  
11 however emphasizes that it is more the vegetation structure determining the macrofauna communities than  
12 that species are typically related to a certain plant species. Descriptions of macroinvertebrate communities  
13 from pond systems should take into account that communities vary considerably depending upon which  
14 habitat types are sampled and that different vegetation forms support different macroinvertebrate  
15 communities.

## 16 17 **6. Conclusions**

18 Within a pond habitat, macroinvertebrate communities differ markedly between sub-habitats (*Open*  
19 *water*, *Bottom substrate*, *Phragmites*, *Nymphaea*, *Callitriche*, *Ceratophyllum*, *Elodea*) in terms of species  
20 abundance composition. Species richness and/or diversity were generally not affected by macrophyte  
21 growth forms, the only exception being a statistically significant difference in mean macroinvertebrate  
22 species diversity between the *Elodea* and *Nymphaea* sub-habitats. Macroinvertebrate relative abundance is  
23 significantly affected by sub-habitat type and whilst the sampling technique used here is semi-quantitative  
24 the differences are so pronounced that they must be considered as significant. Several factors could account  
25 for this difference but these factors are probably all related to availability of microhabitats and food  
26 resources. Those macrophyte sub-habitats yielding the largest number of invertebrates (*Ceratophyllum* and  
27 *Elodea*) were characterised by dense stands of highly branched and dissected plants. This provides huge  
28 relative surface areas for colonization and in addition many refuges for prey species and ambush sites for

1 predatory species. Multivariate analysis showed a clear difference between the macroinvertebrate  
2 assemblages of the macrophyte dominated sites and the *Open water* sub-habitat. The macroinvertebrate  
3 assemblages of the macrophyte dominated sites differ clearly from those of the *Bottom substrate*. Within  
4 the macrophyte dominated sites the macroinvertebrate assemblages show the arrangement of the samples  
5 over a gradient from *Phragmites* and *Nymphaea* via *Callitriche* towards *Elodea* and *Ceratophyllum*. These  
6 macrophyte species represent various growth forms, viz. a helophyte and nymphaeid growth form  
7 (rhizophytes with under water stalks, the latter also with floating leaves), a peplid growth form (rhizophyte  
8 with floating rosettes and oblong or spatulate leaves), a elodeid growth form (submergent rhizophyte with  
9 upright shoots and whorls of linear or oblong leaves) and a ceratophyllid growth form (submerged  
10 pleustophytes with finely divided leaves and without floating leaves).

11 In addition to the above discussion this study clearly demonstrates the importance of macrophyte  
12 presence, and also growth form, on macroinvertebrate species richness and overall abundance within a  
13 small eutrophic pond ecosystem. This therefore highlights the importance of maintaining these freshwater  
14 habitats in a healthy, macrophyte dominated, ecological state, in order to maintain their biodiversity and  
15 conservation value. Furthermore, development of particular macrophyte growth forms in water bodies can  
16 be stimulated by proper management for maintaining a high biodiversity in constructed and semi-natural  
17 water bodies for example urban waters (e.g. Vermonden et al., 2009, 2011).

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## 24 25 **References**

26  
27 Andrews, J.D., Hassler, A.D., 1943. Fluctuations in the animal populations of the littoral zone in Lake  
28 Mendota. Transact. Wisconsin Acad. Sci., Arts and Letters 35, 175-185.

- Clarke, K.R., Gorley, R.N., 2001. Primer v5: User Manual/Tutorial. Primer-E, Plymouth, pp 1-91.
- Cyr, H., Downing, J.A., 1988. The abundance of phytophilous invertebrates on different species of submerged macrophytes. *Freshwat. Biol.* 20, 365-374.
- Den Hartog, C., van der Velde, G., 1988. Structural aspects of aquatic plant communities. In: Symoens, J.J. (ed.) *Vegetation of inland waters. Handbook of vegetation science 15/1*, Kluwer Academic Publishers, Dordrecht, pp 113-153.
- Duggan, I.C., Green, J.D., Thompson, K., Shiel, R.J., 2001. The influence of macrophytes on the spatial distribution of littoral rotifers. *Freshwat. Biol.* 46, 777-786.
- Dvořák, J., Best, E.P.H., 1982. Macro-invertebrate communities associated with the macrophytes of Lake Vechten: structural and functional relationships. *Hydrobiologia* 95, 115-126.
- Furse, M.T., Wright, J.F., Armitage, P.D., Moss, D., 1981. An appraisal of pond-net samples for biological monitoring of lotic macro-invertebrates. *Wat. Res.* 15, 679-689.
- Heck, K.L.J., Orth, R.J., 1980. Seagrass habitats: the roles of habitat complexity, competition and predation in structuring associated fish and motile macroinvertebrate assemblages. In: Kennedy VS (ed) *Estuarine perspectives*. Academic Press, London, 449-464
- Humphries, P., Grouns, J.E., Serafini, L.G., Hawking, J.H., Chick, A.J., Lake, P.S., 1998. Macroinvertebrate sampling methods for lowland Australian rivers. *Hydrobiologia* 364, 209-218
- Jeffries, M., 1993. Invertebrate colonization of artificial pondweeds of differing fractal dimension. *Oikos* 67, 142-148.

- 1
- 2 Jeppesen, E., Søndergaard, M., Christoffersen, K. (eds.), 1998. The structuring role of submerged
- 3 macrophytes in lakes. Ecological Studies 131, 1-423. Springer-Verlag, New York, Berlin, Heidelberg
- 4
- 5 Karassowska, K., Mikulski, J., 1960. Studies of animal aggregations associated with immersed and
- 6 pleustonic vegetations in Lake Druzno. Ekol. polska (Ser. A) 8, 335-353.
- 7
- 8 Khalaf, G., Tachet, H., 1980. Colonisation of artificial substrata by macro-invertebrates in a stream and
- 9 variations according to stone size. Freshwat. Biol. 10, 475-482.
- 10
- 11 Kok, C.J., Hof, C.H.J., Lenssen, J.P.M., van der Velde, G., 1992. The influence of pH on protein and
- 12 phenolics concentration and resource quality of decomposing material of *Nymphaea alba* L.
- 13 (Nymphaeaceae) for the detritivore *Asellus aquaticus* (L.). Oecologia 91, 229-234.
- 14
- 15 Korinkova, J., 1971. Quantitative relations between submerged macrophytes and populations of
- 16 invertebrates in a carp pond. Hydrobiologia 12, 377-382.
- 17
- 18 Kreckler, F.H., 1939. A comparative study of the animal population of certain submerged aquatic plants.
- 19 Ecology 20, 553-562.
- 20
- 21 Marcus, J.H., Sutcliffe, D.W., Willoughby, G., 1978. Feeding and growth of *Asellus aquaticus* (Isopoda) on
- 22 food items from the littoral of Windermere including green leaves of *Elodea*. Freshwat. Biol. 8, 505-519.
- 23
- 24 McAbendroth, L., Ramsay, P.M., Foggo, A., Rundle, S.D., Bilton, D.T., 2005. Does macrophyte fractal
- 25 complexity drive invertebrate diversity, biomass and body size distributions? Oikos 111, 279-290.
- 26
- 27 Mikheev, V.N., Mikheev, A.V., Pasternak, A.F., Valtonen, E.T., 2000. Light mediated host searching
- 28 strategies in a fish ectoparasite, *Argulus foliaceus* L. (Crustacea: Branchiura). Parasitology 120, 409-416.

- 1  
2 Rennie, M.D., Jackson, L.J., 2005. The influence of habitat complexity on littoral invertebrate distributions:  
3 patterns differ in shallow prairie lakes with and without fish. *Can. J. Fish. Aquat. Sci.* 62, 2088-2099.  
4  
5 Reynoldson, T.B., Young, J.O., 1963. The food of four species of lake-dwelling triclads. *J. Anim. Ecol.* 32,  
6 175-191.  
7  
8 Rooke, B., 1986. Macroinvertebrates associated with macrophytes and plastic imitations in the Eramosa  
9 River, Ontario, Canada. *Arch. Hydrobiol.* 106, 307-325.  
10  
11 Rosine, W.N., 1955. The distribution of invertebrates on submerged aquatic plant surfaces in Muskee Lake,  
12 Colorado. *Ecology* 36, 308-314.  
13  
14 Scheffer, M., Achterberg, A.A., Beltman, B., 1984. Distribution of macro-invertebrates in a ditch in  
15 relation to the vegetation. *Freshwat. Biol.* 14, 367-370.  
16  
17 Sheldon, A., Haick, R., 1981. Habitat selection and association of stream insects: a multivariate analysis.  
18 *Freshwat. Biol.* 11, 395-403.  
19  
20 Smolders, A.J.P., Vergeer, L.H.T., van der Velde, G., Roelofs, J.G.M., 2000. Phenolic contents of  
21 submerged, emergent and floating leaves of (semi-)aquatic macrophyte species. Why do they differ? *Oikos*  
22 91, 307-310.  
23  
24 Soszka, J., 1975. Ecological relations between invertebrates and submerged macrophytes in the lake littoral.  
25 *Ekol. polska* 23, 393-415.  
26  
27 Taniguchi, H., Nakano, S., Tokeshi, M., 2003. Influences of habitat complexity on the diversity and  
28 abundance of epiphytic invertebrates on plants. *Freshwat. Biol.* 48, 718-728.



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Ter Braak C.J.F., Smilauer, P., 1998. CANOCO Reference Manual and User's Guide to Canoco for Windows: Software for Canonical Community Ordination (version 4). Microcomputer Power (Ithaca, NY USA). 352 pp.

Tolkamp, H., Both, J., 1978. The organism-substrate relationship in a small Dutch lowland stream. Preliminary results. Verhandl. Internat. Verein. Limnol. 20, 1509-1515.

Van der Valk, A.G., 2006. The biology of freshwater wetlands. Oxford University Press, 173 pp.

Van der Velde, G., 1980. Studies in nymphaeid-dominated systems. PhD thesis University of Nijmegen, 169 pp.

Vermonden, K., Leuven, R.S.E.W., van der Velde, G., van Katwijk, M.M., Roelofs, J.G.M., Hendriks, A.J., 2009. Urban drainage systems: An undervalued habitat for aquatic macroinvertebrates. Biol. Conserv. 142, 1105-1115.

Vermonden, K., van der Velde, G., Leuven, R.S.E.W., 2011. Key factors for biodiversity of surface waters in climate proof cities. Resources, Conserv. Recycling. Doi:10.1016/j.resconrec.2011.01.003.

Warfe, D.M., Barmuta, L.A., 2004. Habitat structural complexity mediates the foraging success of multiple predator species. Oecologia 141, 171-178.

1 **Table 1.** Description of the seven habitat types sampled in Rossall pond including depth  
2 range at which the habitat types were found and/or sampled. Growth forms of  
3 macrophytes according to the classification by Den Hartog and Van der Velde (1988).

Habitat	Description	Complexity	Depth range (cm) sampled
Open water	Open water with no macrophytes present.	Highly simple	50-120
Bottom substrate	Bottom sediment consisting mainly of fine gravel and silt. No macrophytes present.	Complex	30-100
Elodeid	Predominantly <i>Elodea canadensis</i> . Submerged rhizophyte with upright shoots and with small but broad, oval leaves typically in whorls of 4 at each node. No floating leaves.	Moderately complex	40-100
Ceratophyllid	Predominantly <i>Ceratophyllum demersum</i> . Submerged rhizophyte or pleustophyte with whorls of 5-12 leaves at each node. Leaves typically forked once or twice. No floating leaves.	Highly complex	40-100
Peplid	Predominantly <i>Callitriche</i> sp. Caulescent rhizophyte with branched structure with elliptical floating leaves in a rosette and linear submerged leaves.	Moderately complex	40-60
Nymphaeid	Predominantly <i>Nymphaea alba</i> . Large, floating, shield-shaped leaves attached to long, simple, submerged stalks.	Simple	70-120
Helophyte	Predominantly <i>Phragmites australis</i> although frequently interspersed with individuals of <i>Typha latifolia</i> and <i>Iris pseudacorus</i> . Plants rooting in the bottom, with basal	Simple	30-50

parts continuously  
submerged running  
vertically through water  
column; leaves and  
inflorescences far above the  
water surface.

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- 1 **Table 2.** Full species list including number of individuals sampled for each of the seven  
2 habitats in Rossall pond and functional feeding groups: **D** ó detritivores; **SGS** ó  
3 shredders, grazers and scrapers; **F** ó filtrators; **PrC** ó predatory carnivores; **PaC** ó  
4 parasitic carnivores.

<b>Habitat</b> <b>Taxa</b>	Feeding group	<i>Phragmites</i> <i>australis</i>	<i>Nymphaea</i> <i>alba</i>	<i>Callitriche</i> sp.	<i>Ceratophyllum</i> <i>demersum</i>	<i>Elodea</i> <i>canadensis</i>	<i>Substrate</i>	<i>Open</i> <i>water</i>
<b>Insecta</b>								
<b>Diptera</b>								
<i>Chironomidae</i>	<b>D</b>	2	14	10	21	10	32	
<i>Ceratopogonidae</i>	<b>D</b>	1						
<i>Pedicia</i> sp.	<b>D</b>	1						
<i>Dixa</i> sp.	<b>F</b>	1		1				
<i>Eristalis</i> sp.	<b>D</b>	2		7			5	2
<b>Odonata</b>								
<i>Ischnura elegans</i>	<b>PrC</b>	29	14	29	61	29	45	
<i>Enallagma cyathigerum</i>	<b>PrC</b>	12	31	23	3	15	10	1
<i>Aeshna grandis</i>	<b>PrC</b>						8	
<b>Coleoptera</b>								
<i>Hygrotus inaequalis</i>	<b>PrC</b>			3				
<i>Hyphydrus ovatus</i>	<b>PrC</b>		4				2	
<i>Dryops</i> sp.	<b>PrC</b>							1
<b>Heteroptera</b>								
<i>Notonecta</i> sp.	<b>PrC</b>	3		2		1	2	
<i>Corixa punctata</i>	<b>D</b>	37	4	19	9	13	67	2
<b>Ephemeroptera</b>								
<i>Cloeon dipterum</i>	<b>SGS</b>	1	3					
<b>Trichoptera</b>								
<i>Arthripsodes aterrimus</i>	<b>SGS</b>						2	
<b>Lepidoptera</b>								
<i>Elophila nymphaeata</i>	<b>SGS</b>		3					
<b>Megaloptera</b>								
<i>Sialis lutaria</i>	<b>PrC</b>			1	1		64	
<b>Crustacea</b>								
<i>Eurycercus lamellatus</i>	<b>F</b>	16		9	11	5	11	1
<i>Simocephalus vetulus</i>	<b>F</b>	2	2			2		
<i>Asellus aquaticus</i>	<b>SGS</b>	12	5	111	276	204	163	1
<i>Argulus foliaceus</i>	<b>PaC</b>	1	2				2	8
<i>Cypris</i> sp.	<b>F</b>	1		11			14	
<i>Crangonyx pseudogracilis</i>	<b>SGS</b>	2	4	8		7	1	
<b>Mollusca</b>								
<b>Bivalvia</b>								
<i>Musculium lacustre</i>	<b>F</b>	2						
<i>Sphaerium corneum</i>	<b>F</b>						61	

<i>Pisidium</i> sp.	<b>F</b>		<b>1</b>	<b>1</b>			<b>1</b>	
<b>Gastropoda</b>								
<i>Physa fontinalis</i>	<b>SGS</b>	<b>5</b>			<b>10</b>	<b>1</b>	<b>7</b>	<b>2</b>
<i>Segmentina complanata</i>	<b>SGS</b>	<b>9</b>	<b>5</b>	<b>5</b>		<b>8</b>	<b>3</b>	
<i>Radix peregra/ovata</i>	<b>SGS</b>	<b>6</b>	<b>3</b>		<b>1</b>	<b>1</b>		
<i>Planorbis carinatus</i>	<b>SGS</b>	<b>7</b>		<b>10</b>		<b>7</b>	<b>4</b>	
<i>Planorbarius comeus</i>	<b>SGS</b>				<b>24</b>	<b>5</b>		
<i>Radix auricularia</i>	<b>SGS</b>		<b>8</b>				<b>5</b>	
<b>Hirudinea</b>								
<i>Helobdella stagnalis</i>	<b>PrC</b>	<b>1</b>					<b>2</b>	
<i>Hemiclepsis marginata</i>	<b>PaC</b>				<b>7</b>			
<i>Glossiphonia complanata</i>	<b>PrC</b>		<b>3</b>		<b>4</b>			
<b>Turbellaria</b>								
<i>Dugesia lugubris</i>	<b>PrC</b>	<b>8</b>	<b>16</b>	<b>71</b>	<b>357</b>	<b>425</b>	<b>9</b>	<b>1</b>
<b>Oligochaeta</b>								
<i>Tubifex</i> sp.	<b>D</b>						<b>5</b>	
<b>Arachnidae</b>								
<i>Argyroneta aquatica</i>	<b>PrC</b>	<b>2</b>						
Hydracarina	<b>PrC</b>		<b>3</b>	<b>5</b>	<b>10</b>	<b>10</b>	<b>5</b>	

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**Table 3.** Macroinvertebrate species richness, abundance and BPD (Species diversity) values for the seven different habitats. Also shown is the proportion of the total pond species richness found in each of the habitats (% of total richness). % of total abundance = the proportion of the total number of macroinvertebrates sampled that were obtained from each habitat. The most abundant species for each habitat type are also given. Numbers in parentheses = 1 standard error.

	<i>Phragmites</i>	<i>Nymphaea</i>	<i>Callitriche</i>	<i>Ceratophyllum</i>	<i>Elodea</i>	<i>Substrate</i>	<i>Open water</i>
Richness	25	20	18	14	17	26	9
% of total richness	61	48.8	44	34.1	41.5	63.4	22
Number of unique taxa	3	1	1	1	0	3	1
Mean richness per sample (n = 10)	7 (2)	7.1 (2.2)	9 (2.3)	7.1 (1.7)	8.2 (1.8)	11.5 (3.3)	1.6 (1.2)
Abundance	167	127	326	789	753	530	19
% of total abundance	6.2	4.7	12	29.1	27.8	19.6	0.7
Most abundant taxa	<i>Corixa punctata</i>	<i>Enallagma cyathigerum</i>	<i>Asellus aquaticus</i>	<i>Dugesia lugubris</i>	<i>Dugesia lugubris</i>	<i>Asellus aquaticus</i>	<i>Argulus foliaceus</i>
Mean abundance per sample (n = 10)	16.7 (6.2)	32.6 (15.8)	78.9 (39.0)	12.7 (4.2)	53 (22.9)	1.9 (1.4)	75.3 (24.8)
Species Diversity	0.22	0.24	0.34	0.45	0.56	0.31	0.42
Mean diversity per sample (n = 10)	0.38 (0.12)	0.34 (0.13)	0.4 (0.14)	0.51 (0.09)	0.36 (0.18)	0.67 (0.37)	0.56 (0.11)

1 **Table 4.** Analyses of similarity (ANOSIM) test results (corresponding to the nMDS plot  
2 of Fig. 3) indicating significant differences between communities of the different habitat  
3 types. R-values for pair-wise comparisons varying between 0 and 1, indicating the degree  
4 of separation (from R=0; communities completely overlap, to R=1; communities are  
5 completely separated) are shown when differences are significant ( $\alpha = 0.0024$  after  
6 Bonferroni correction; ns = not significant).

	<i>Phragmites</i>	<i>Nymphaea</i>	<i>Callitriche</i>	<i>Ceratophyllum</i>	<i>Elodea</i>	<i>Substrate</i>	<i>Open water</i>
<i>Phragmites</i>							
	Overall comparison of communities: p=0.001, R=0.517						
<i>Nymphaea</i>	0.410						
<i>Callitriche</i>	ns	0.485					
<i>Ceratophyllum</i>	0.565	0.776	0.684				
<i>Elodea</i>	0.432	0.592	0.315	ns			
<i>Substrate</i>	0.535	0.752	0.574	0.925	0.870		
<i>Open water</i>	0.466	0.567	0.566	0.571	0.577	0.539	

## Figure captions

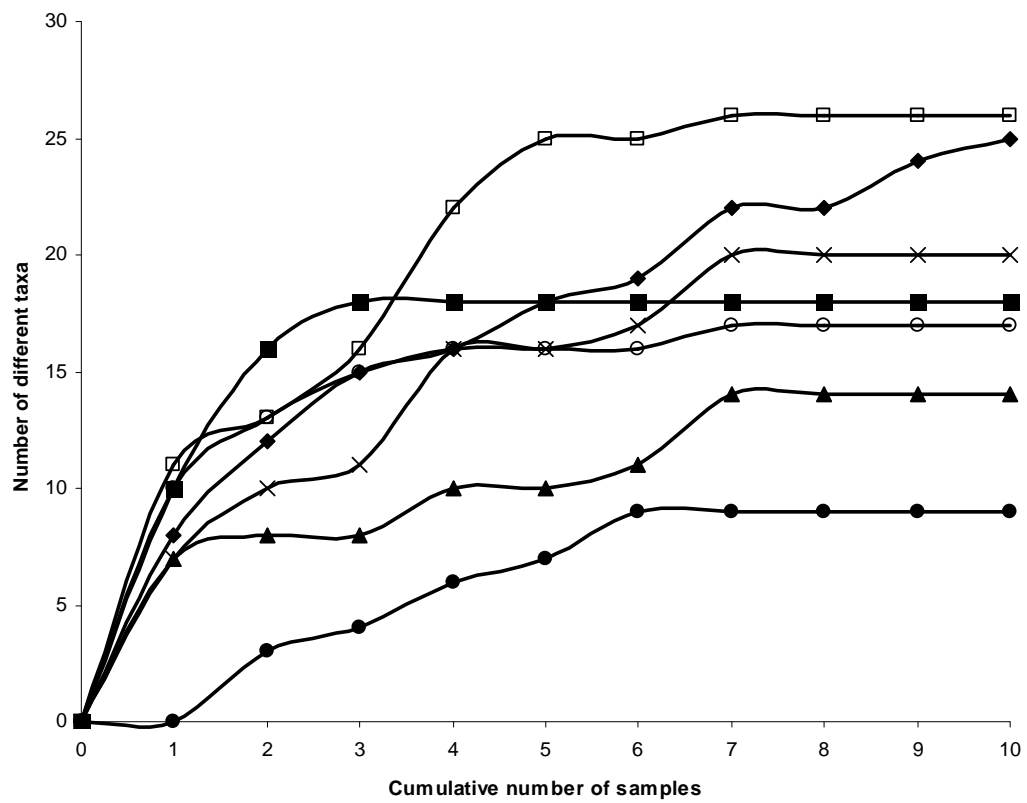
**Fig. 1.** Taxon accretion curves for all seven habitats. ○ = Open water habitat; □ = Bottom substrate habitat; ◆ = *Phragmites* habitat; × = *Nymphaea* habitat; ■ = *Callitriche* habitat; ● = *Elodea* habitat ; ▲ = *Ceratophyllum* habitat.

**Fig. 2.** CCA analysis without (a) and with (b) Open water habitat data. Chironomidae (1); Ceratopogonidae (2); *Pedicia* sp. (3); *Dixa* sp. (4); *Eristalis* sp. (5); *Ischnura elegans* (6); *Enallagma cyathigerum* (7); *Aeshna grandis* (8); *Hygrotus inaequalis* (9); *Hyphydrus ovatus* (10); *Dryops* sp. (11); *Notonecta* sp. (12); *Corixa punctata* (13); *Cloeon dipterum* (14); *Arthripsodes aterrimus* (15); *Elophila nymphaeata* (16); *Sialis lutaria* (17); *Eurycercus lamellatus* (18); *Simocephalus vetulus* (19); *Asellus aquaticus* (20); *Argulus foliaceus* (21); *Cypris* sp. (22); *Crangonyx pseudogracilis* (23); *Musculium lacustre* (24); *Sphaerium corneum* (25); *Pisidium* sp. (26); *Physa fontinalis* (27); *Segmentina complanata* (28); *Radix peregra* (29); *Planorbis carinatus* (30); *Planorbarius corneus* (31); *Radix auricularia* (32); *Helobdella stagnalis* (33); *Hemiclepsis marginata* (34); *Glossiphonia complanata* (35); *Dugesia lugubris* (36); *Tubifex* sp. (37); *Argyroneta aquatica* (38); Hydracarina (39). Insert = CCA analysis with *Open water* data included.

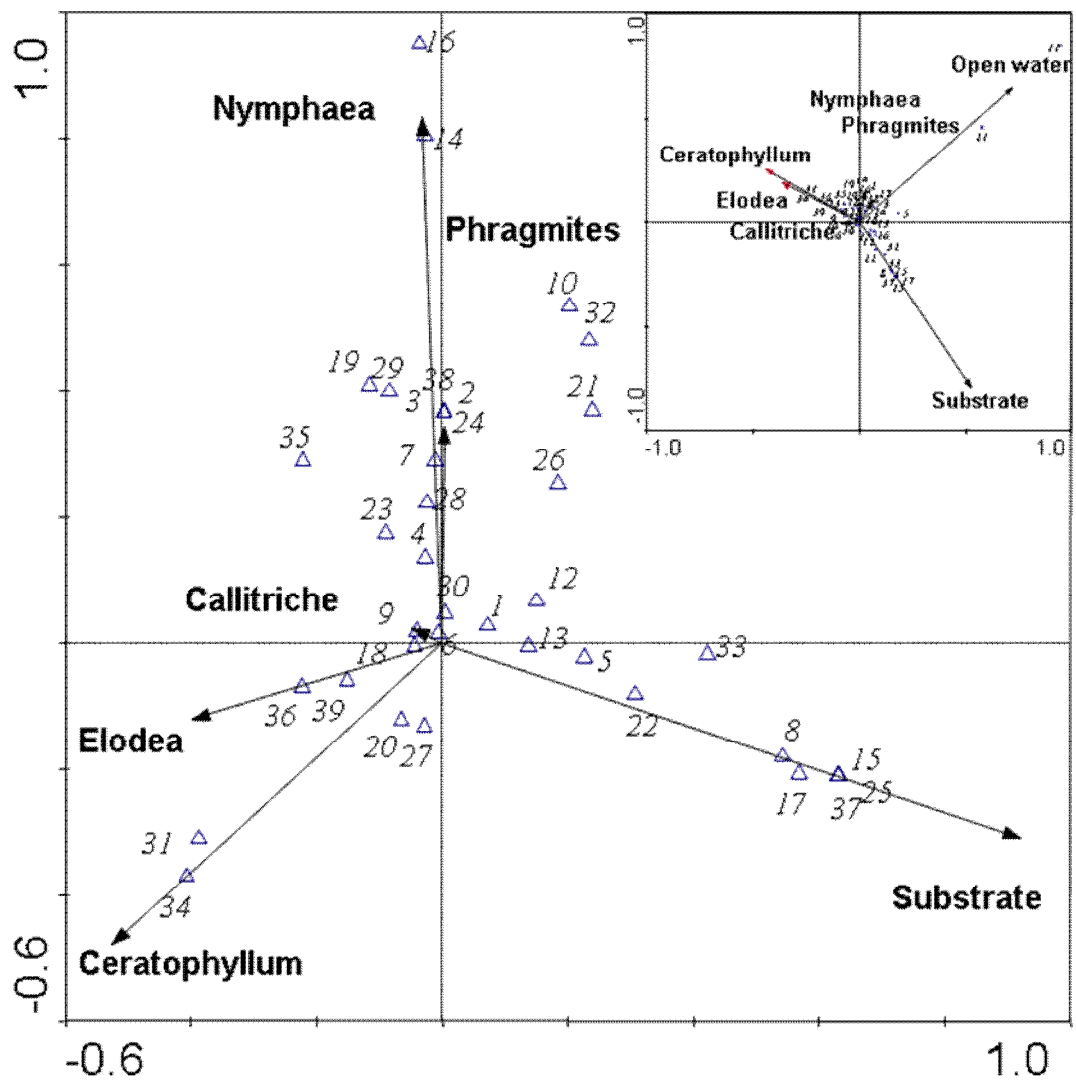
**Fig. 3.** Non-metric multi-dimensional scaling (nMDS) plot indicating the similarity of macrofauna communities per habitat type, showing the individual samples.



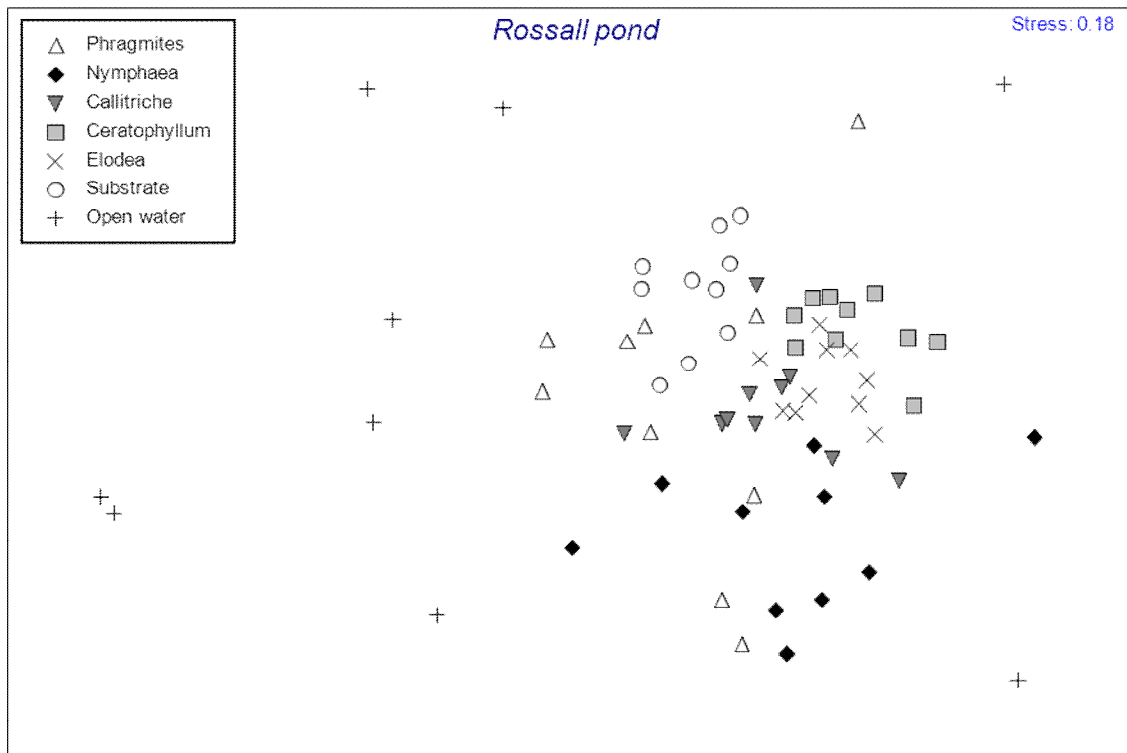
1 **Fig. 4.** Proportion of different macroinvertebrate feeding groups in Rossall pond showing  
2 proportions for seven different habitats also. **D** ó detritivores; **SGS** ó shredders, grazers  
3 and scrapers; **F** ó filter feeders; **PrC** ó predatory carnivores; **PaC** ó parasitic carnivores.  
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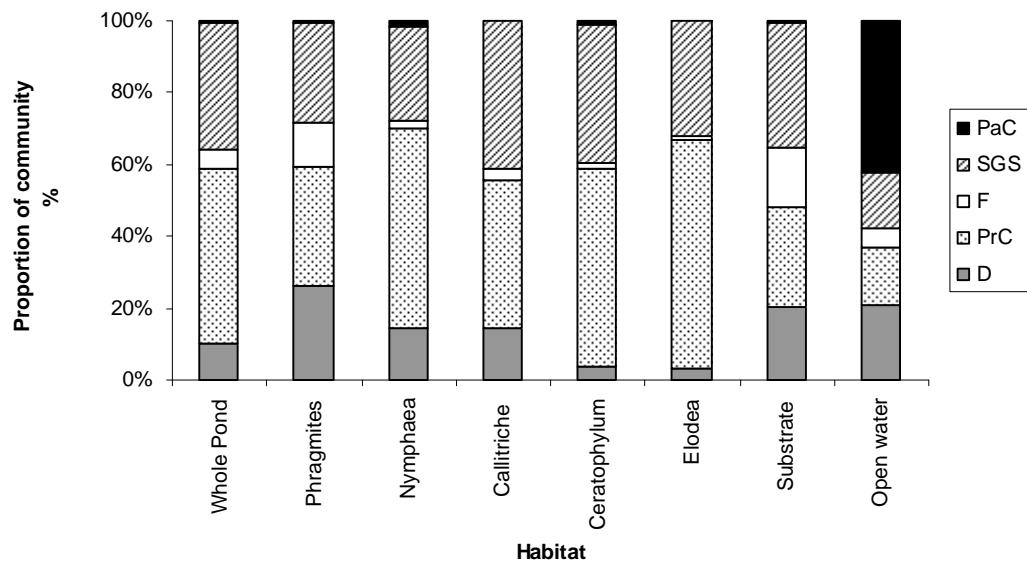
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2 **Fig. 1.**  
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**Fig. 3.**



**Fig. 4**